

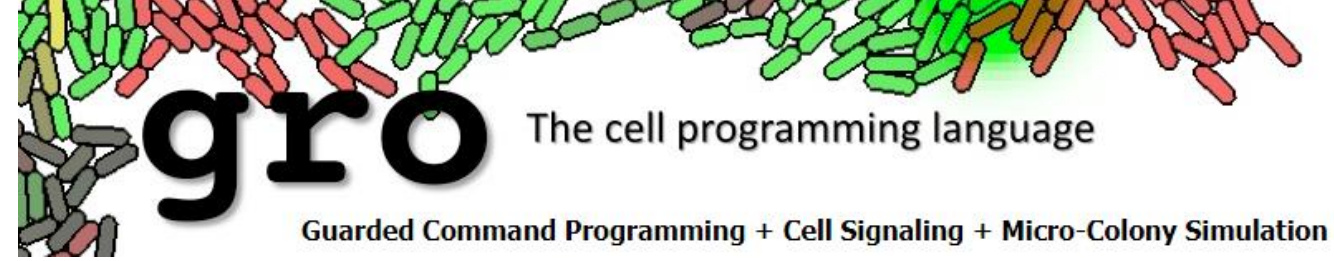
Gro Programming and Simulation:

BE 240 Lecture 5

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05/07/2020

What is gro

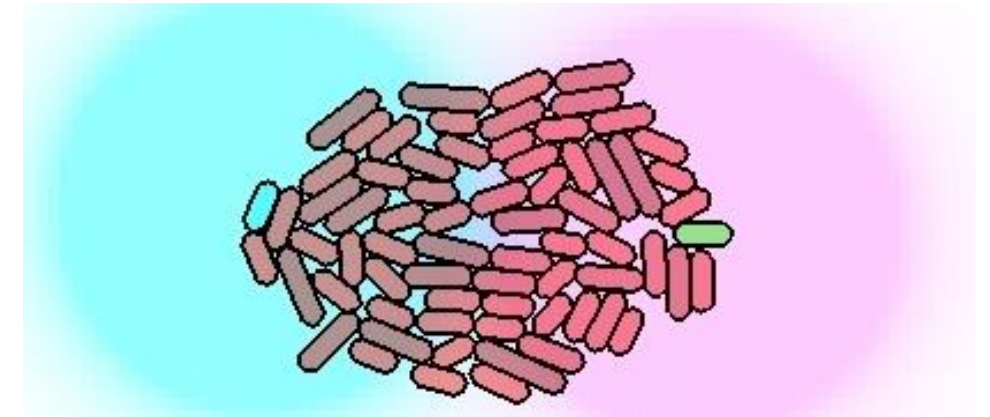


Developed by [The Klavins Lab](http://depts.washington.edu/soslab/gro/index.html), University Washington, Seattle, WA

<http://depts.washington.edu/soslab/gro/index.html>

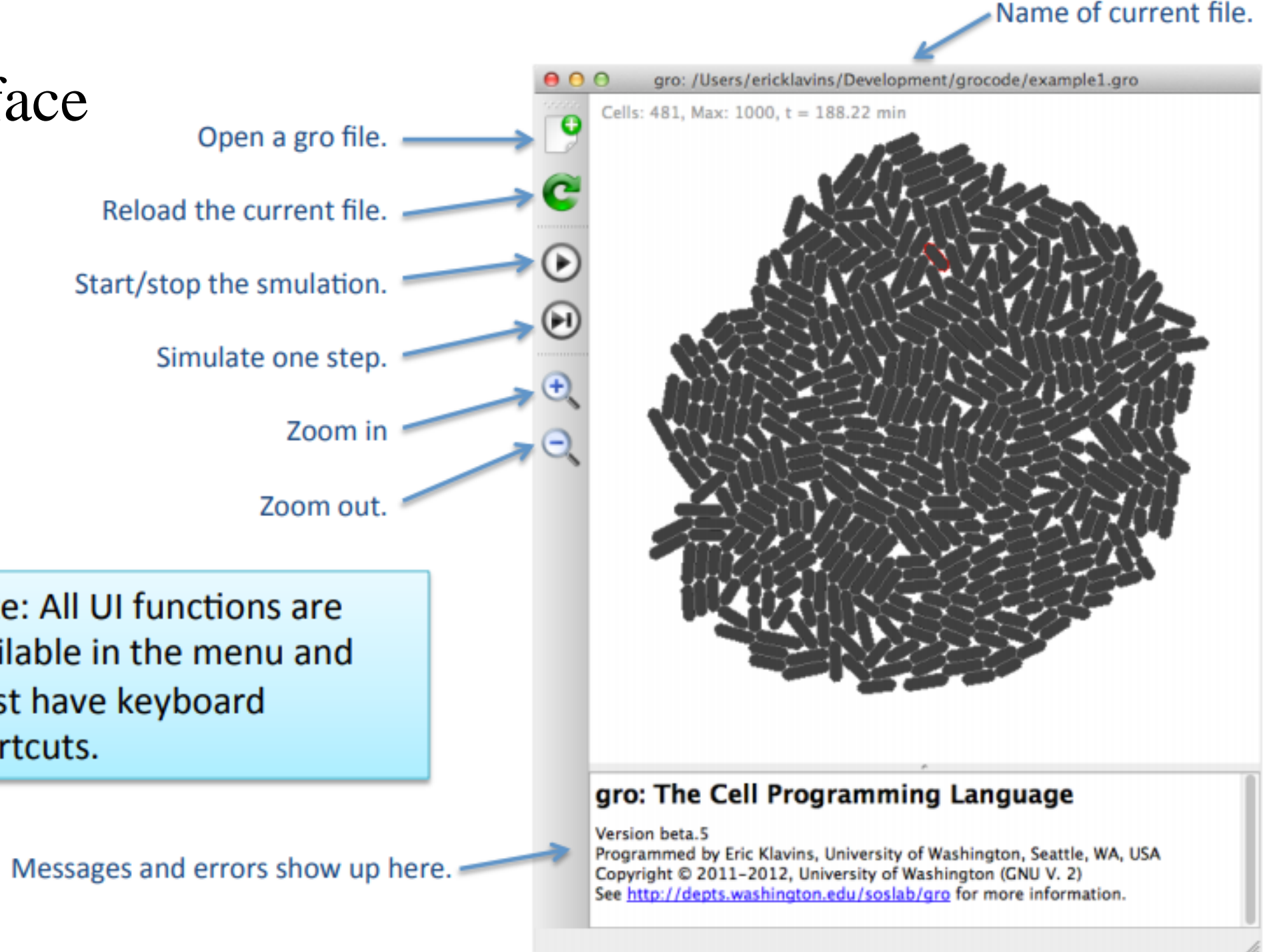
A tool for programming, modeling, specifying and simulating of **multicellular** behaviors in **growing** microcolonies in **a 2D environment**.

- Cell growth/division
- Cell crowding
- Signal diffusion
- Molecular reactions



E. coli microcolonies growing in a single layer \leftrightarrow fluorescence microscope

User interface



Installation

<http://depts.washington.edu/soslab/gro/download.php>

Mac OS X

Date	Release	Description
Aug. 23, 2012	gro_mac_beta.4.dmg	This version has an improved gui, reacting signals, more modifiable parameters, new examples, and the return of the command line. See the changelog below for more details. Note: gro needs to stay in the same directory as the examples and include folders that are included in the disk image.
Older	gro_mac_beta.3.dmg gro_a.5.4.tar.gz	

small bug: path configuration

Windows 7

Date	Release	Description
Aug. 23, 2012	gro_win_beta.4.zip	This version is (hopefully) the same as the mac version above, except compiled for Windows. Qt supposedly takes care of cross-compatability issues. Since the development team (i.e. Eric) uses a Mac to test everything, the Windows version might stil have some issues -- which you should please report if you find any.
Older	gro_win_beta.3.zip gro_win_a.4.tar.gz	

Installation

<https://github.com/murrayrm/gro>

MacOS

You will need the following packages in order to compile gro:

- CCL: <https://github.com/klavinslab/ccl>
- Chipmunk 5.3.5: <https://chipmunk-physics.net/release/Chipmunk-5.x/>
- XCode

Once you have these pre-requisties, you can install `gro` by running `qmake` and telling it where to find the ccl and chipmunk source directories (which should also have the compiled library files).

```
qmake CCL=<cclpath> CHIPMUNK=<chipmunkpath>  
make
```

This will create directory `gro.app` that can run using the command

```
open gro.app
```

Installation

<https://github.com/murrayrm/gro>

Linux

You will need the following packages in order to compile gro:

- CCL: <https://github.com/klavinslab/ccl>
- Chipmunk 5.3.5: <https://chipmunk-physics.net/release/Chipmunk-5.x/>
- Linux build tools (via apt): build-essential, flex, bison, libreadline-dev
- OpenGL: freeglut3-dev

Once you have these pre-requisites, you can install `gro` by running `qmake` and telling it where to find the `ccl` and `chipmunk` source directories (which should also have the compiled library files).

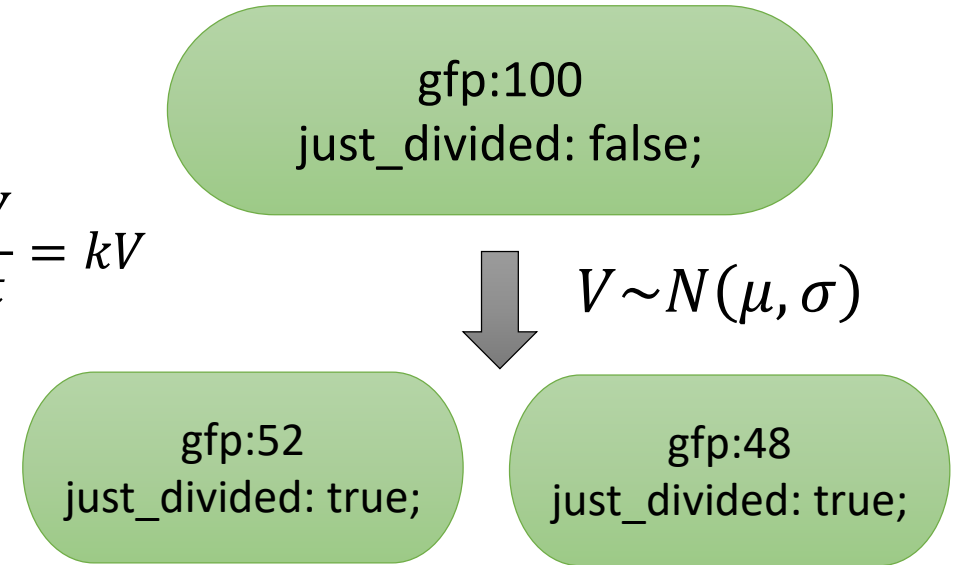
```
qmake CCL=<cclpath> CHIPMUNK=<chipmunkpath>  
make
```

The file `useful/chipmunk.gro` in the main `gro` directory is available to allow compilation of chipmunk via `qmake`. To use it, copy `useful/chipmunk.gro` to the `chipmunk` main source directory and run `qmake` then `make`.

Simulation environment

E. coli-like bacteria

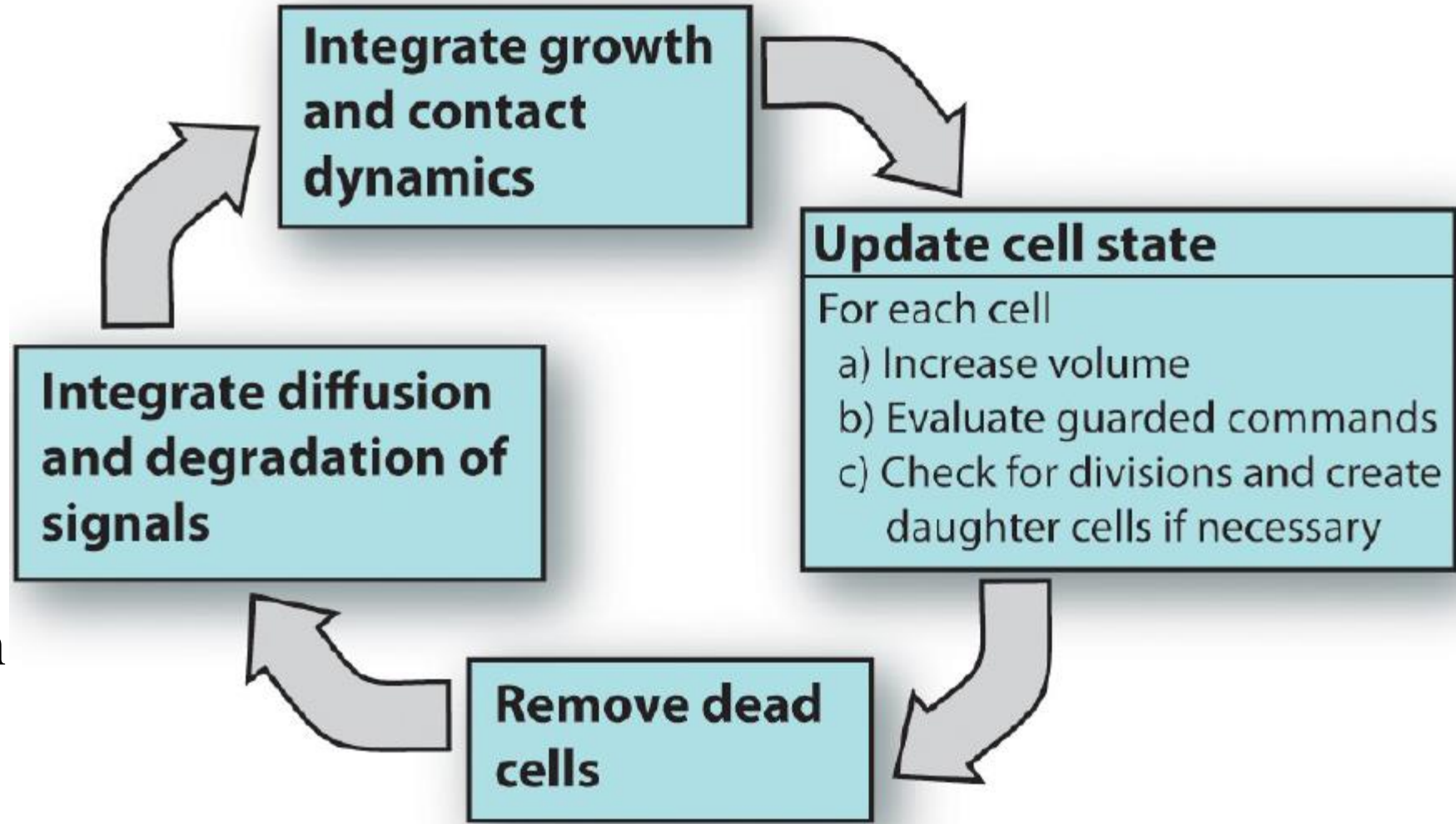
- Cell growth: volume grows exponentially $\frac{dV}{dt} = kV$
- Cell division: divides after doubled in size
- Cell death: removed after dead
- Cell crowding: physics contact (Chipmunk)
- Signal diffusion: 2D grid of square elements (Finite element method)
- Molecular reactions: stochastic events (guarded command g:c)
- Chemostat mode



Simulation environment

E. coli-like bacteria

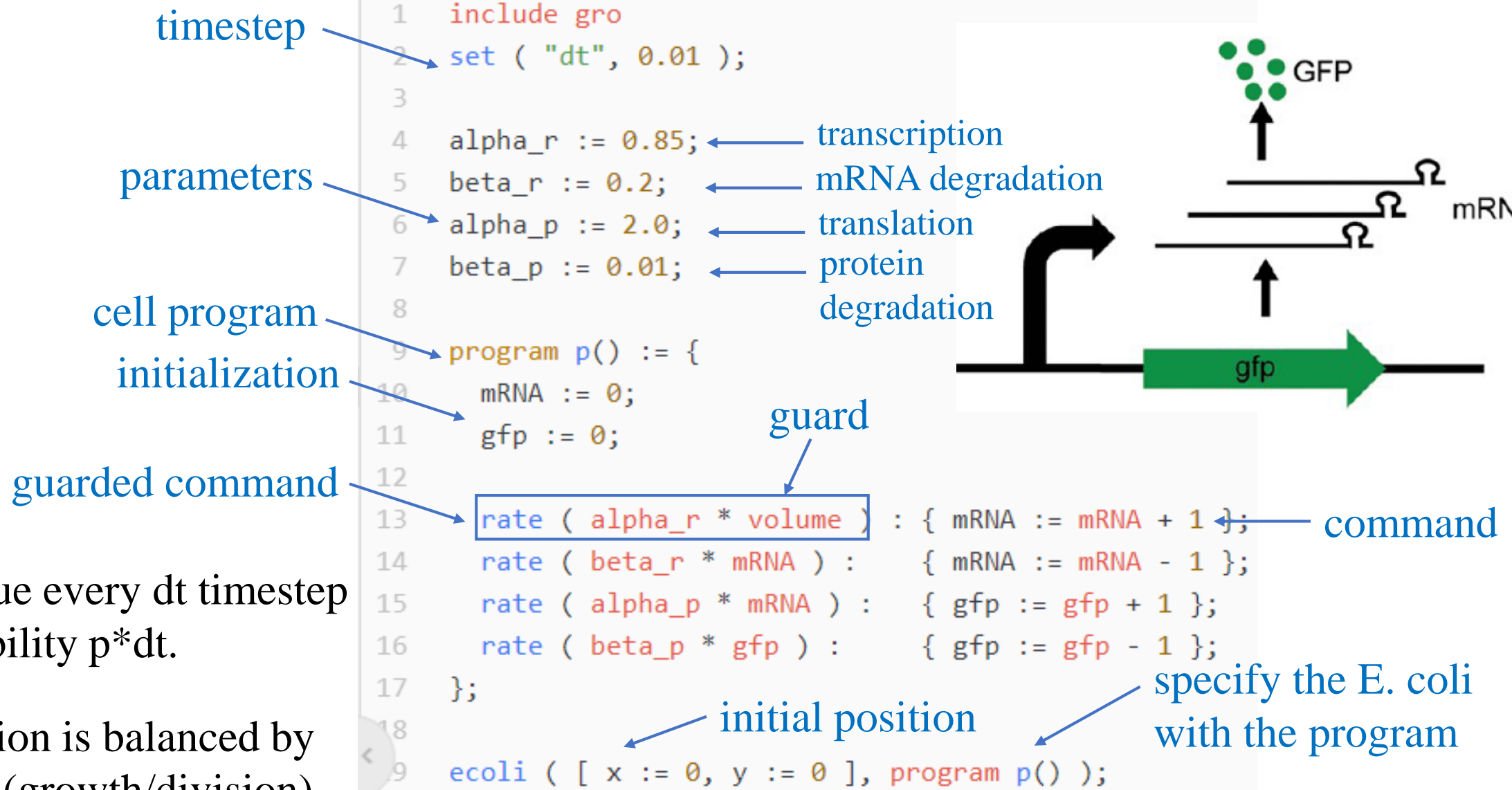
- Cell growth
- Cell division
- Cell death
- Cell crowding
- Signal diffusion
- Molecular reaction



gro programming: inside the cell

example_gfp_simple.gro

Program p ()



gro programming: more functions

Mass action
propensities:

```
rate ( alpha_r * volume ) : { mRNA := mRNA + 1 };
rate ( beta_r * mRNA ) : { mRNA := mRNA - 1 };
rate ( alpha_p * mRNA ) : { gfp := gfp + 1 };
rate ( beta_p * gfp ) : { gfp := gfp - 1 };
```

Define
functions:

$$f_{hill}(v, k, x) = v \frac{x}{k + x}$$

```
fun hill v k x . v * x / ( k + x );
```

Non-mass action
propensities:

```
rate ( hill v k x ) : { gfp := gfp + 1 };
```

$$f_{logistic}(v, kmax, x) = v \left(1 - \frac{x}{kmax}\right) x$$

```
fun logistic v kmax x . v * ( 1 - x / kmax ) * x;
```

Other functions:

$$f_{fact}(n) = n!$$

```
fun fact n .
  if n <= 0
  then 1
  else n * fact (n-1)
end;
```

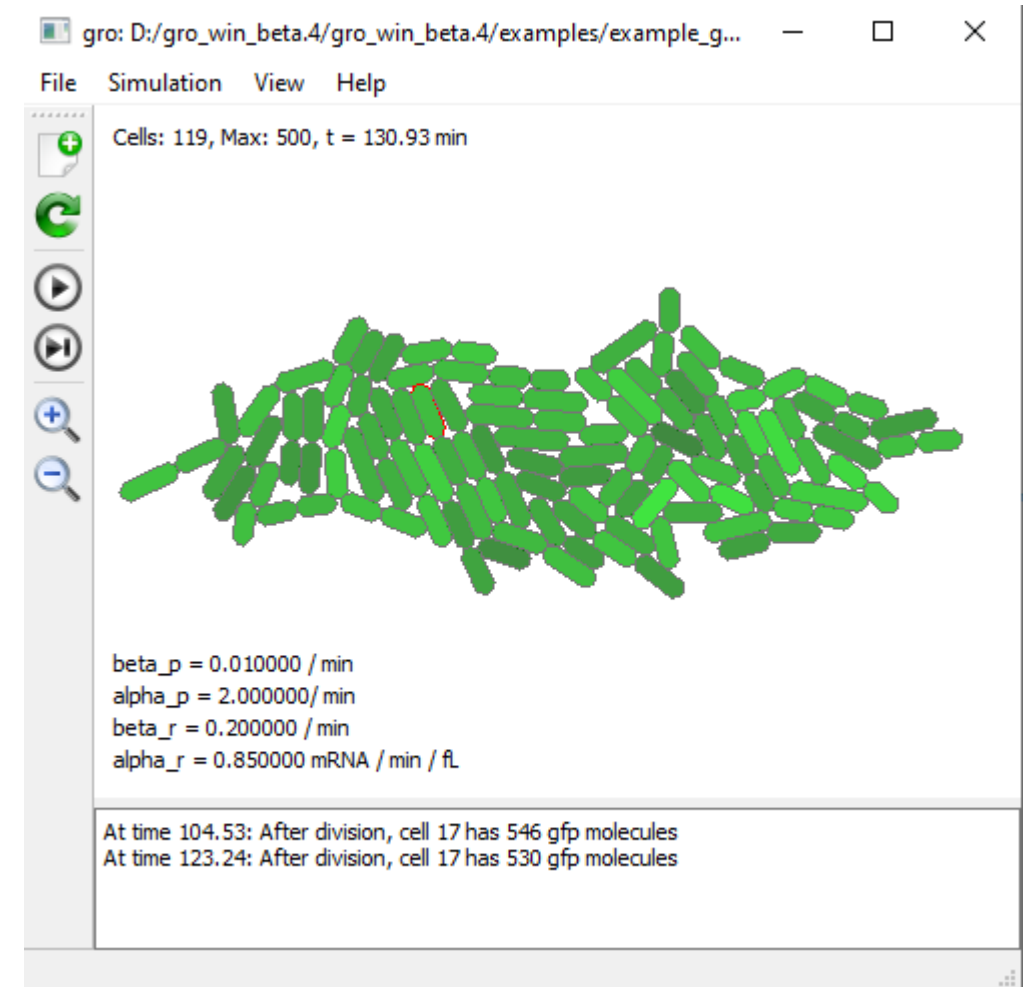
```
+, -, *, ^, /, %
sqrt(x), sin(y)
```

gro simulation `example_gfp_const.gro`

- Keep track of the time

```
9  program p() := {
10    mRNA := 0;
11    gfp := 0;
12
13    rate ( alpha_r * volume ) : { mRNA := mRNA + 1 };
14    rate ( beta_r * mRNA ) :    { mRNA := mRNA - 1 };
15    rate ( alpha_p * mRNA ) :   { gfp := gfp + 1 };
16    rate ( beta_p * gfp ) :     { gfp := gfp - 1 };
17
18    r := [ t := 0 ]; ← hide time in a record
19    selected & just_divided : {
20      print ( "At time ", r.t, ": After division, cell ",
21        id, " has ", gfp, " gfp molecules" )
22    }; ← track the cell id
23    true : { r.t := r.t + dt };
24  };
```

time increments




gro simulation `example_gfp_const.gro`

- Keep track of the time by composing two program

```
2  program report(period) := {  
43    t := 0;  
44    s := 0;  
45    needs gfp; ← get the shared gfp  
46    true : { t := t + dt, s := s + dt }  
47  s >= period : {  
48    print ( id, ", " , t, ", " , gfp, ", " , volume, "\n" ),  
49    s := 0;  
50  }  
51 };  
52  
53 program q() := p() + report(1.0) sharing gfp, mRNA;
```

Only shared variables get cut
in half when cell divides

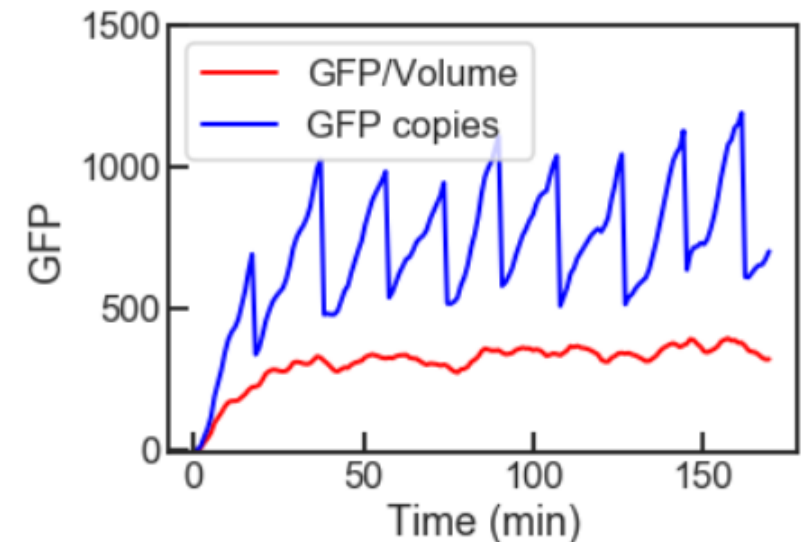
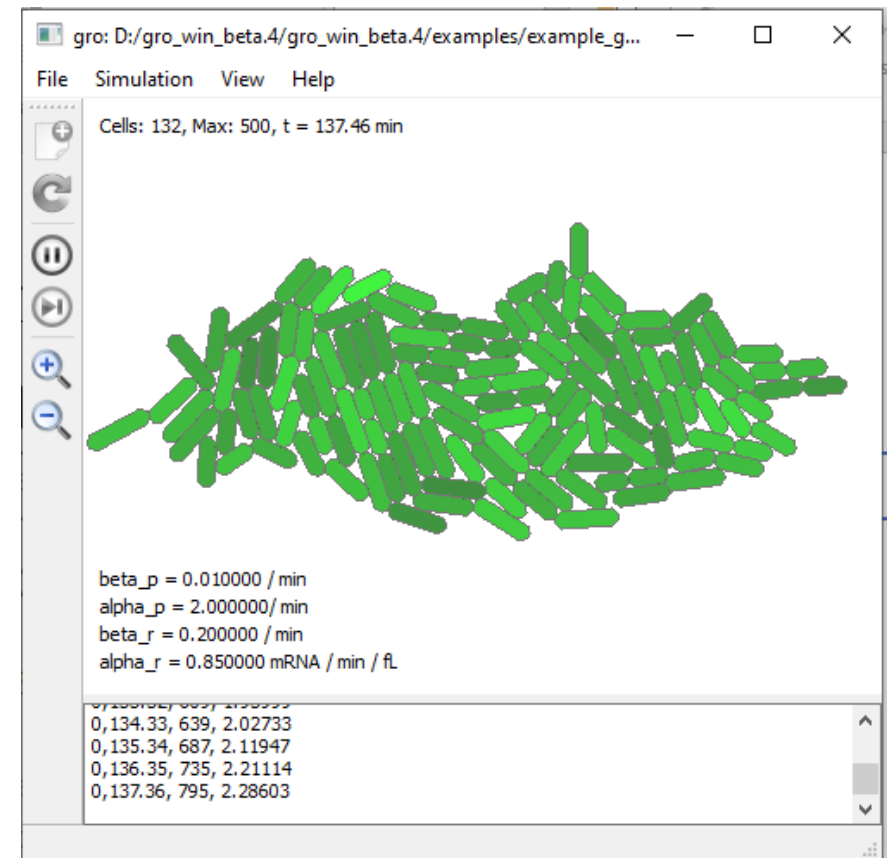


```
9  program p() := {  
10    mRNA := 0;  
11    gfp := 0;  
12  
13    rate ( alpha_r * volume ) : { mRNA := mRNA + 1 };  
14    rate ( beta_r * mRNA ) : { mRNA := mRNA - 1 };  
15    rate ( alpha_p * mRNA ) : { gfp := gfp + 1 };  
16    rate ( beta_p * gfp ) : { gfp := gfp - 1 };  
17  };
```

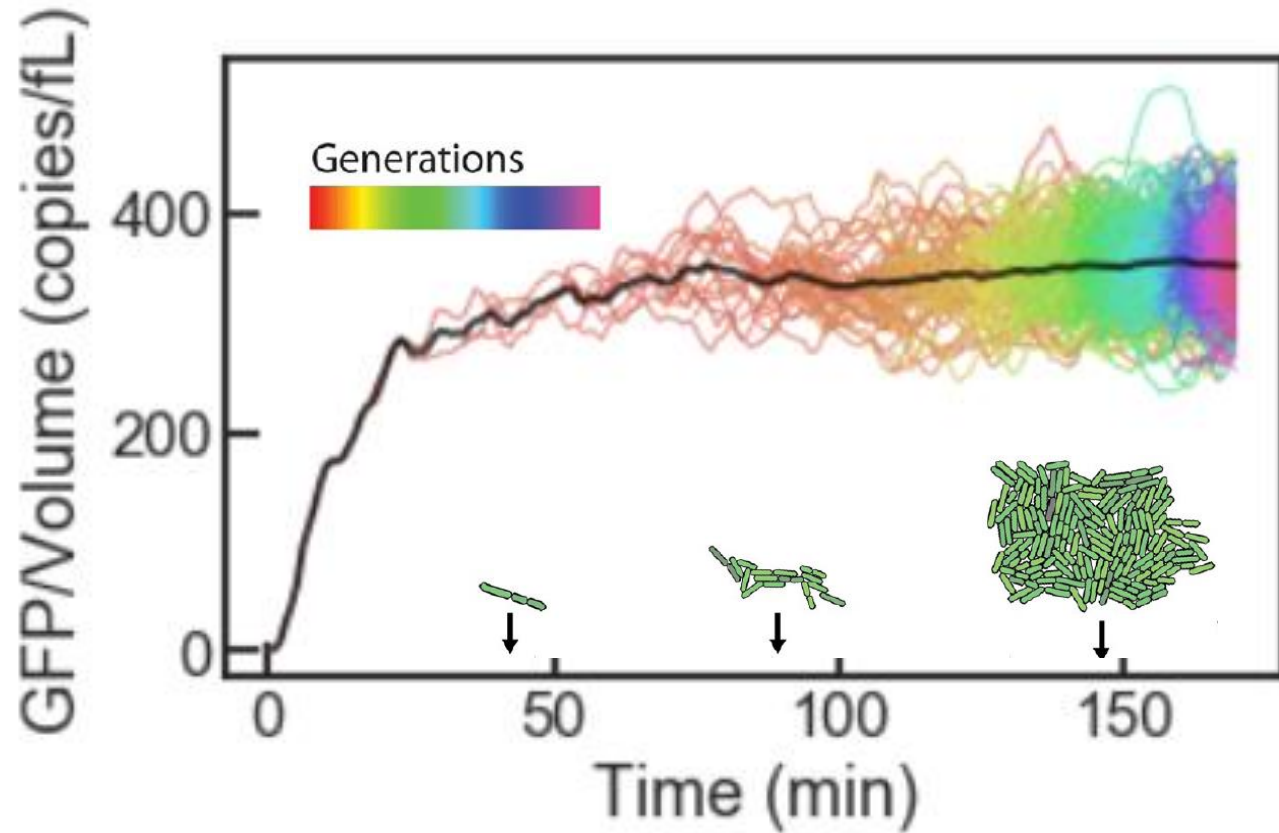
gro simulation `example_gfp_const.gro`

- Save data in csv ← write here

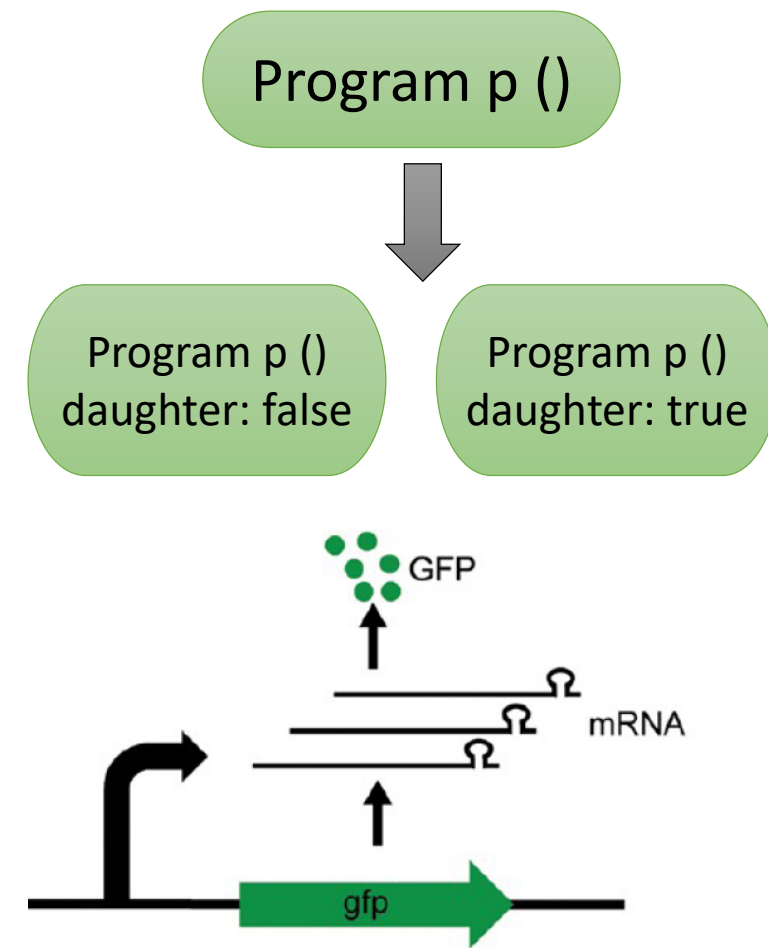
```
9  fp := fopen ( "/tmp/example_gfp.csv", "w" );
10
11  program p() := {
12    mRNA := 0;
13    gfp := 0;
14
15    rate ( alpha_r * volume ) : { mRNA := mRNA + 1 };
16    rate ( beta_r * mRNA ) :    { mRNA := mRNA - 1 };
17    rate ( alpha_p * mRNA ) :  { gfp := gfp + 1 };
18    rate ( beta_p * gfp ) :    { gfp := gfp - 1 };
19
20    r := [ t := 0, s := 0 ]; ← r.t track simulation time
21    id = 0 & r.s >= 1.0 : { ← r.s time gap for saving data
22      print ( id, ",", r.t, ",", gfp, ",", volume, "\n" ),
23      fprintf ( fp, id, ",", r.t, ",", gfp, ",", volume, "\n" ),
24      r.s := 0; ← save data
25    };
26    true : { r.t := r.t + dt, r.s := r.s + dt }; ← time increments
27  };
```



gro simulation `example_gfp_const.gro`



- Cell divides →
- program is copied
 - numerical variables are cut in half approximately
- 2^n copies of program after n generations



gro simulation

example_gfp_const.gro

- Save snapshots for movies

```
45 program main() := {
46   t := 0; // framerate time tracker
47   s := 0; // total time tracker
48   n := 0;
49   true : { t := t + dt, s := s + dt }
50   t > 5 : {
51     snapshot ( "/movie/expression" <> if n <10 then "0" else "" end <> toString(n) <> ".tif" ),
52     n := n + 1,
53     t := 0
54   }
55   s > 150 : { stop() }
56 };
```

File path and name

expressionNN.tif

takes snapshots every 5
program minutes and stops
at 150 minutes

gro simulation example_gfp_const.gro

- Chemostat Mode

```
1 include gro
2 chemostat( true );
3 set ( "chemostat_width", 40 );
4 set ( "chemostat_height", 200 );
```

← set chemostat size

Chemostat mode allows you to keep a more or less fixed population of cells in the simulation.

- Set population cap

```
2 set ( "population_max", 50);
```

Cells: 50, Max: 50, t = 108.52 min

Cell = 50

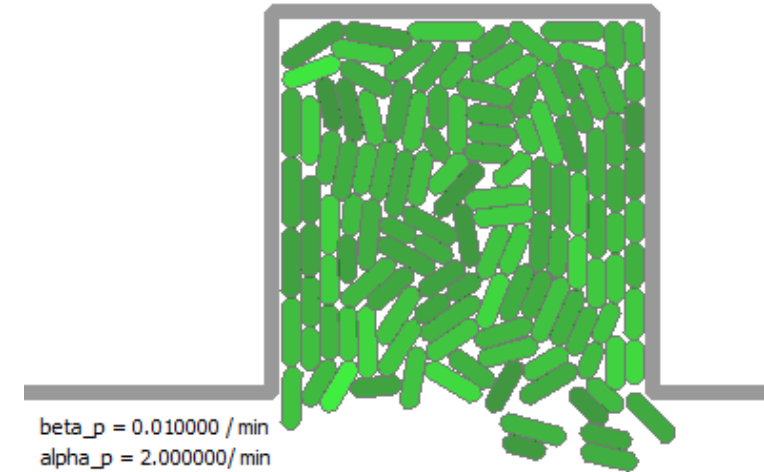


beta_p = 0.010000 / min
alpha_p = 2.000000 / min
beta_r = 0.200000 / min
alpha_r = 0.850000 mRNA / min / fL

Population limit reached. Increase the population limit via the Simulation menu, or by setting the parameter "population_max" in your gro program.

Cells: 130, Max: 500, t = 136.24 min

Cell = 130

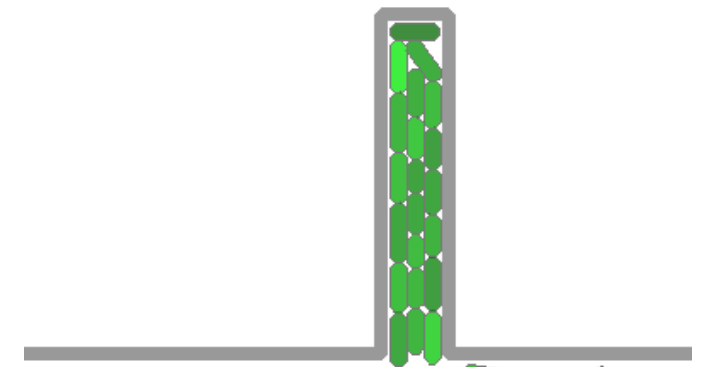


beta_p = 0.010000 / min
alpha_p = 2.000000 / min
beta_r = 0.200000 / min
alpha_r = 0.850000 mRNA / min / fL

Media flow →

Cells: 24, Max: 500, t = 198.82 min

Cell = 24

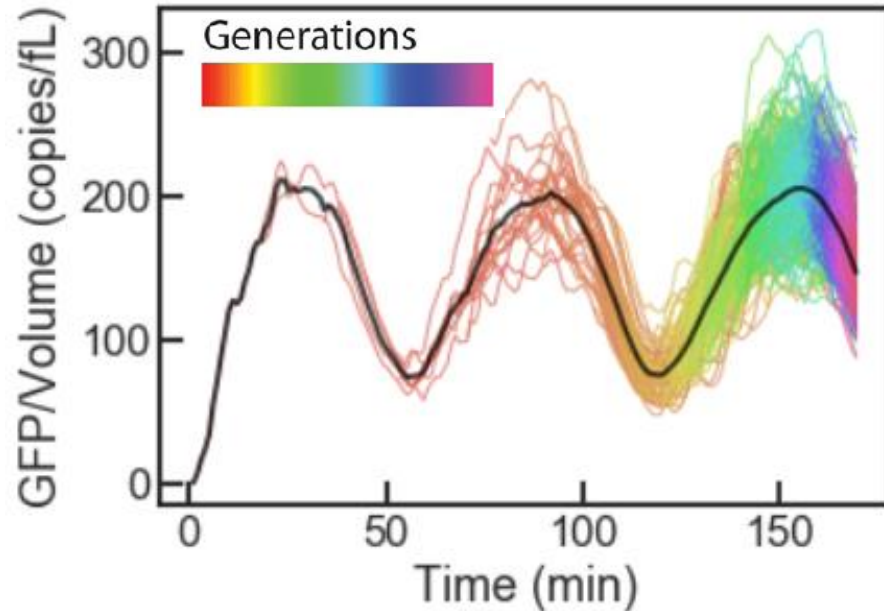
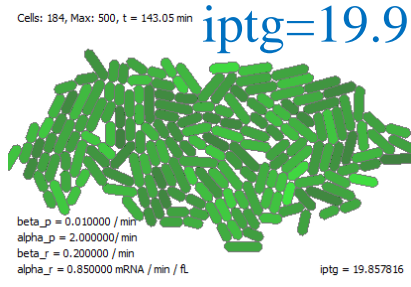


beta_p = 0.010000 / min
alpha_p = 2.000000 / min
beta_r = 0.200000 / min
alpha_r = 0.850000 mRNA / min / fL

Media flow →

example_gfp_sin.gro gro simulation

- Global control



Cells: 57, Max: 500, t = 112.25 min

iptg=0.26



beta_p = 0.010000 / min
alpha_p = 2.000000 / min
beta_r = 0.200000 / min
alpha_r = 0.850000 mRNA / min / fL

iptg = 0.262028

```

9  fun hill v k x . v * x / ( k + x );
10 alpha := 1.0;
11 K := 10.0;
12 iptg := 1.0;
13
14 fp := fopen ( "/tmp/example_gfp.csv", "w" );
15
16 program p() := {
17   mRNA := 0;
18   gfp := 0;
19
20   rate ( alpha_r * volume * hill alpha K iptg ) : { mRNA := mRNA + 1 };
21   rate ( beta_r * mRNA ) : { mRNA := mRNA - 1 };
22   rate ( alpha_p * mRNA ) : { gfp := gfp + 1 };
23   rate ( beta_p * gfp ) : { gfp := gfp - 1 };
24 };
25
26 ecoli ( [ x := 0, y := 0 ], program p() );
27
28 program main() := {
29   t := 0;
30   true : {
31     t := t + dt,
32     iptg := 50 * ( 1 + sin(0.1*t) ),
33     clear_messages ( 1 ),
34     message ( 1, "iptg = " <> toString(iptg) ) }
35 };

```

global variable

iptg induced transcription

global variable control (not associated with a cell)

change iptg according to a sine wave

gro simulation

- Simulation control

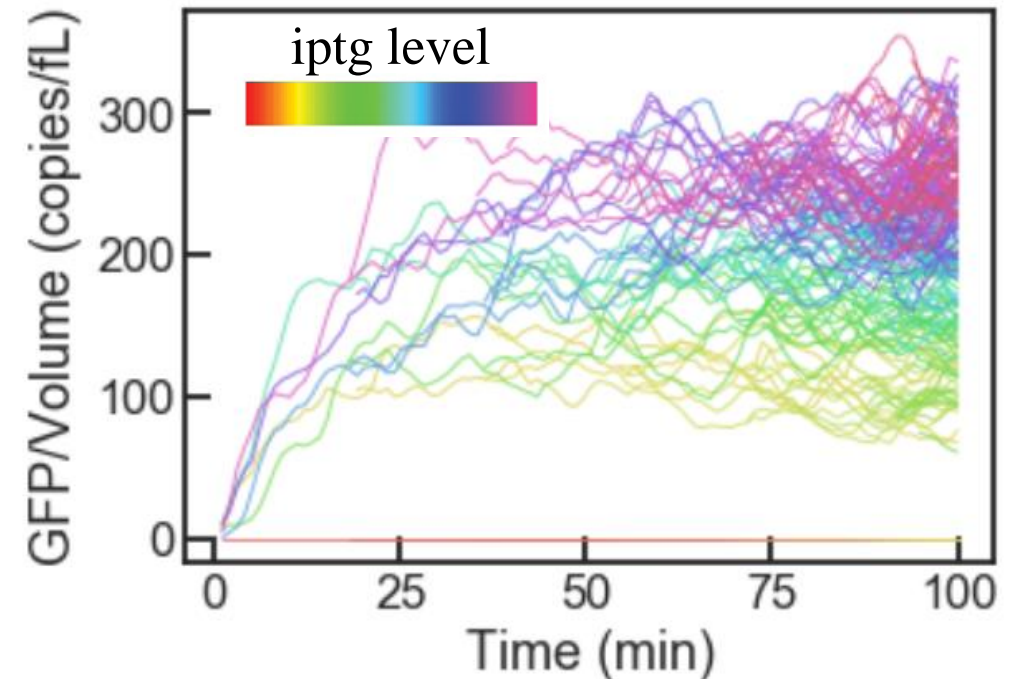
Run multiple simulations with different iptg.

```
46 program main() := {  
47   t := 0;  
48   true : { t := t + dt }  
49   t > 100 : {  
50     print ( iptg, ", ", maptocells gfp/volume end ),  
51     iptg := iptg + 5, ← change iptg level every  
52     reset(),          100 min  
53     ecoli ( [ x := 0, y := 0 ], program p() ),  
54     start(),  
55     t := 0  
56   }  
57   iptg > 30 : { stop() }  
58 };
```

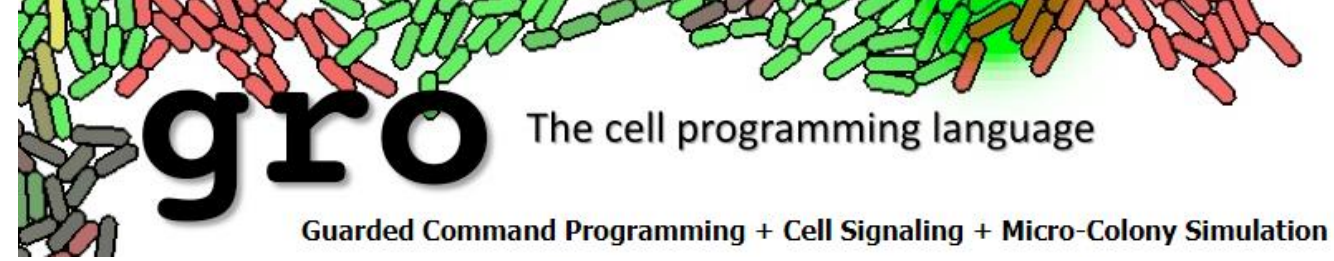
reset() deletes all the cells, so you need to add a new cell after calling it

start(), stop(), and reset()

```
16 program p() := {  
17   mRNA := 0;  
18   gfp := 0;  
19  
20   rate ( alpha_r * volume * hill alpha K iptg ) : { mRNA := mRNA + 1 };  
21   rate ( beta_r * mRNA ) : { mRNA := mRNA - 1 };  
22   rate ( alpha_p * mRNA ) : { gfp := gfp + 1 };  
23   rate ( beta_p * gfp ) : { gfp := gfp - 1 };  
24 };  
25  
26 ecoli ( [ x := 0, y := 0 ], program p() );
```



What is gro



A tool for the simulation of **multicellular** behaviors in **a 2D environment**.

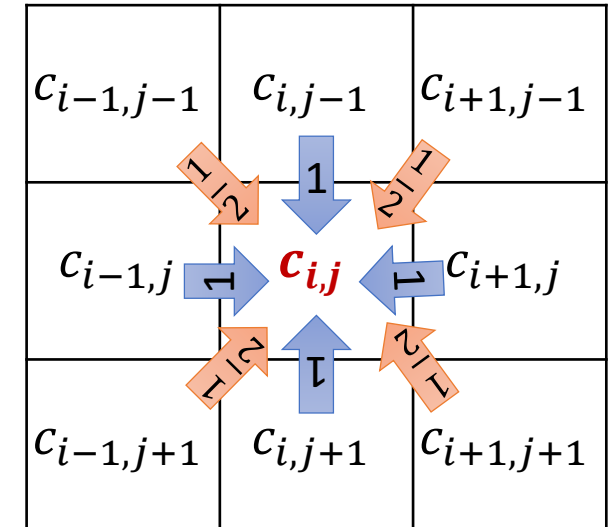
- **Cell growth/division : try with bioscrape lineage**
- **Cell crowding**
- **Signal diffusion**
- **Molecular reactions (with noise) : try with bioscrape**

Spatial pattern formation, heterogeneity in populations...

gro signals : finite element model

- The 2D environment of the simulation is gridded up into $0.5 \mu\text{m} * 0.5 \mu\text{m}$ elements.
- The concentration of each element $c_{i,j}$ is updated at each time step via

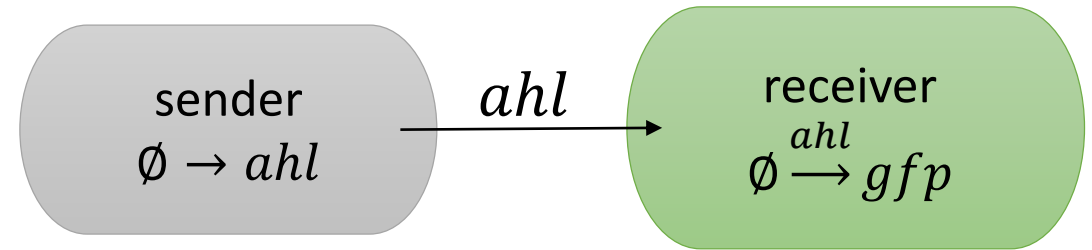
$$\begin{aligned}\Delta c_{i,j} \\ &= k_{diff} (0.5c_{i+1,j-1} + c_{i+1,j} \\ &\quad + 0.5c_{i+1,j+1} + c_{i,j-1} + c_{i,j+1})\end{aligned}$$



- Unspecified height, the units for signal concentration are proportional to moles / L.
- Euler integration: high diffusion rate of a signal \rightarrow obvious numerical errors \rightarrow decrease dt

gro programming: signals

- Sender - receiver



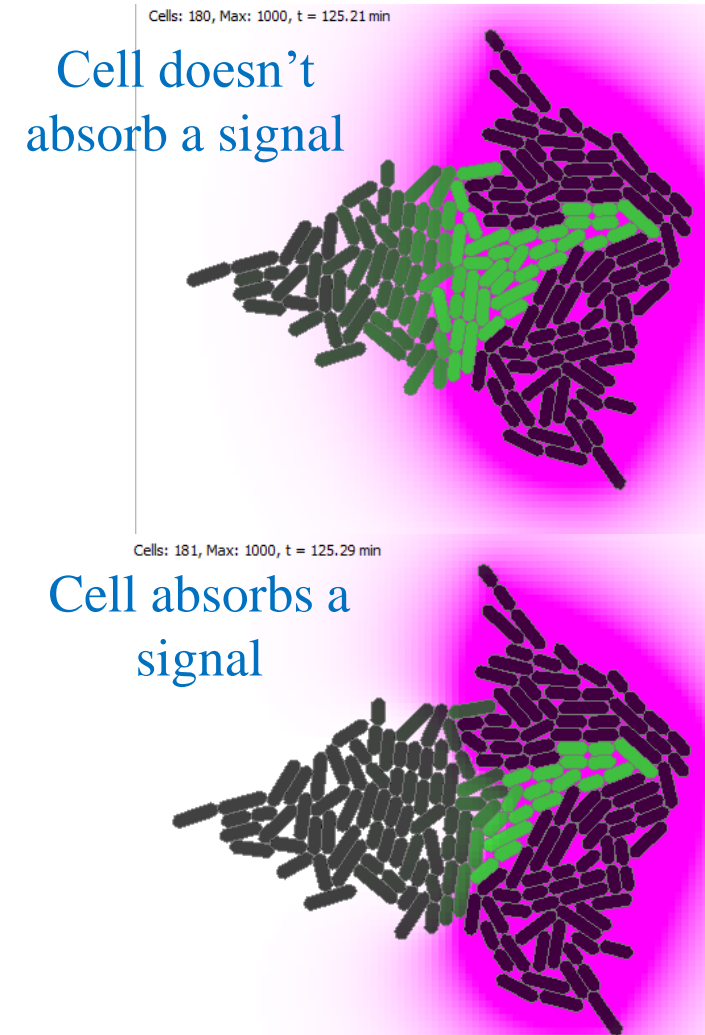
```
1 include gro
2 ahl := signal(5, 0.1);
3 k := 2;
4
5 program signaler() := {
6   true : { emit_signal(ahl, 0.2) }
7 };
8
9 program receiver() := {
10  gfp := 0;
11  rate( k * get_signal(ahl) ) : { gfp := gfp + 1 }
12  true : { absorb_signal(ahl, 0.1) }
13 };
14
15 ecoli ( [x:=50, theta:=3.14/2], program signaler() );
16 ecoli ( [x:=-50], program receiver() );
```

declare a signal
(diffusion, degradation)

send signal with
certain amount of ahl

Cell senses a signal

Cell absorbs a signal (signal
removal, e.g. nutrients)

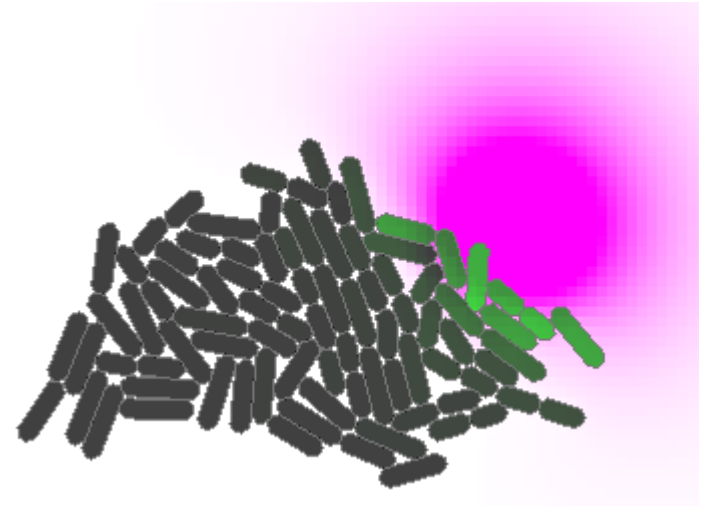


gro programming: signals

- Environment signals

```
15 program main() := {  
16   true : { set_signal(ahl,50,-50,10) }  
17 };  
18  
19 ecoli ( [x:=-50], program receiver() );
```

position and amount of
signal in the environment



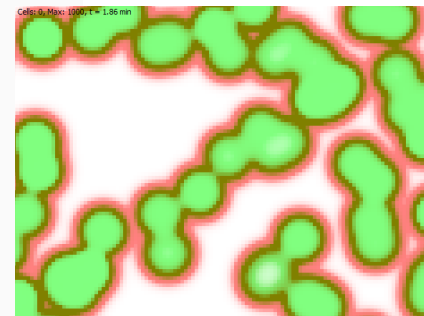
- Reaction-diffusion

chemicals that can (1) react with each other (or themselves) and (2) diffuse

```
28 X := signal ( 1.0, 0.0 );  
29 Y := signal ( 1.0, 0.0 );  
30  
31 reaction ( {X,Y}, {Y,Y}, 5 );  
32 reaction ( {X}, {X,X}, 5 );  
33 reaction ( {Y}, {}, 5 );  
34  
35 foreach i in range 100 do {  
36   set_signal ( X, rand(800)-400, rand(800)-400, 1 ),  
37   set_signal ( Y, rand(800)-400, rand(800)-400, 1 )  
38 } end;
```

signal interacting reactions
(reactants, products, rate)

signal initialization



gro programming: signals

- Bioprocessing

```
4 biomass := signal(0, 0);
5 enzyme := signal(4, 0.3);
6 food := signal(5, 0.1);
7
8 reaction({biomass, enzyme}, {food, enzyme}, 5);
9 set("ecoli_growth_rate", 0.0);
10
11 program bioprocessor() := {
12   true : {
13     set("ecoli_growth_rate", get_signal(food));
14     emit_signal(enzyme, 1);
15   }
16 };
17
18 program main() := {
19   t := 0;
20   true: { t := t + dt }
21   foreach i in range 500 do {
22     set_signal(biomass, rand(300), (rand(500)-250), 10)
23   } end;
24 };
25 ecoli ( [], program bioprocessor() );
```

declare a signals (no diffusion or degradation for biomass)

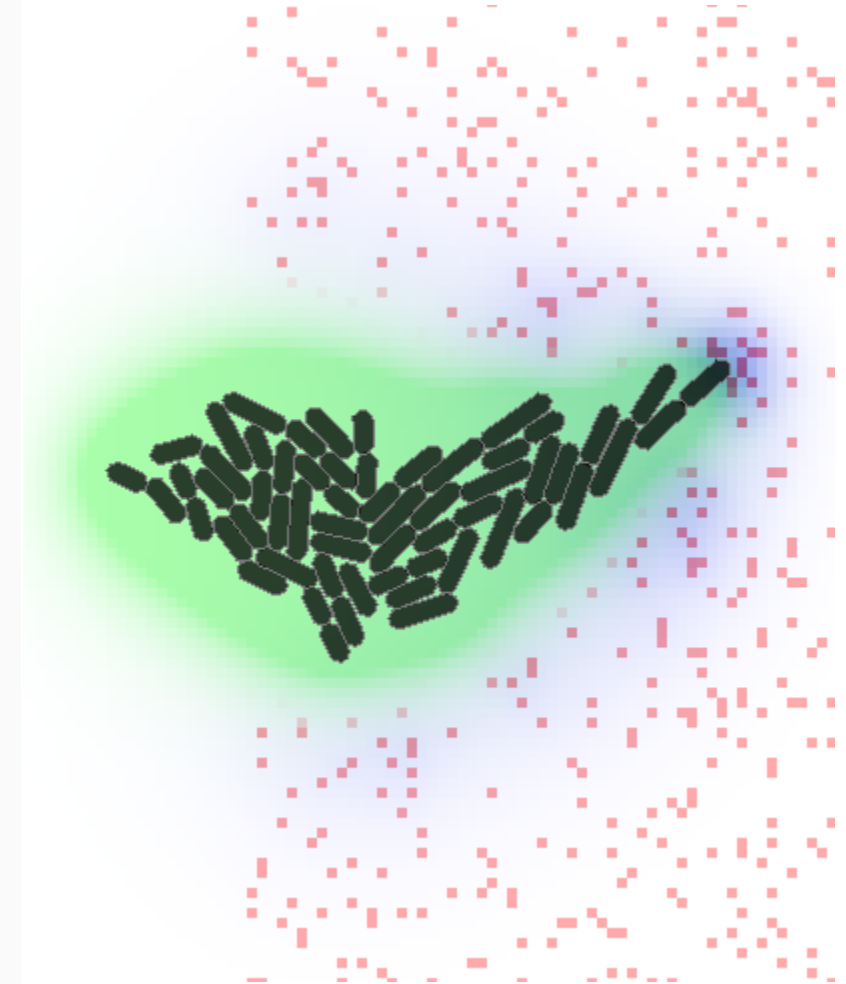
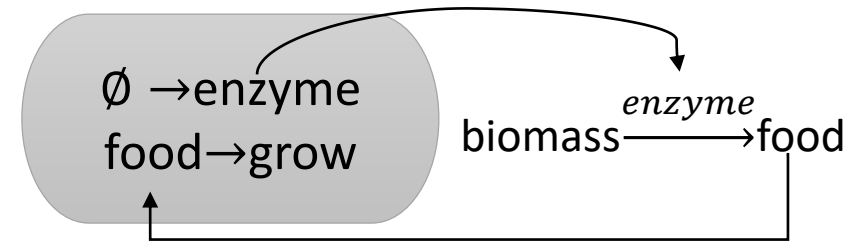
enzyme catalyzes biomass and generates food

set cell growth rate without food

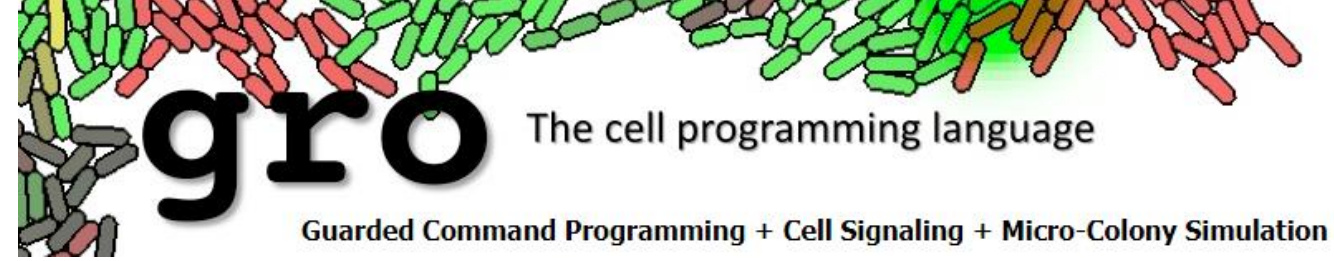
cell growth depends on food

cell secretes enzymes

initializing biomass



What is gro



A tool for the simulation of **multicellular** behaviors in **a 2D environment**.

- **Cell growth/division : try with bioscrape lineage**
- **Cell crowding**
- **Signal diffusion : try with PDE solver**
- **Molecular reactions (with noise) : try with bioscrape**

The environment shapes the microbial organisms.

Cells self-organize into certain patterns via signaling.

Try simulate the same circuit/system in well-mixed (bioscrape) VS spatial (gro).