

Alliance for Cellular Signaling

- 40 laboratories investigating basic questions in cell signaling
 - How complex is signal processing in cells?
 - What is the structure and dynamics of the network?
 - Can functional modules be defined?
 - Key Advantage of AfCS: High quality data from single cell type
 - All findings will be available to signaling community (www.signaling-gateway.org)
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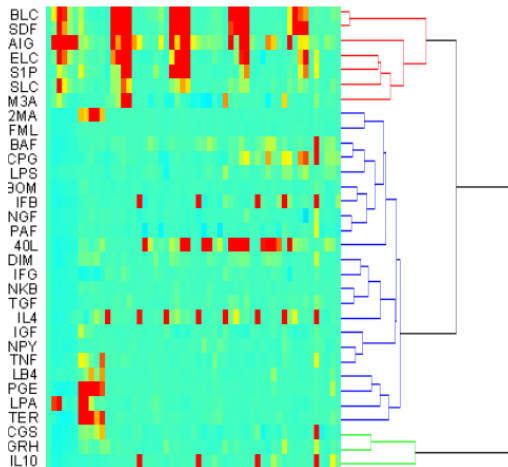
AfCS data



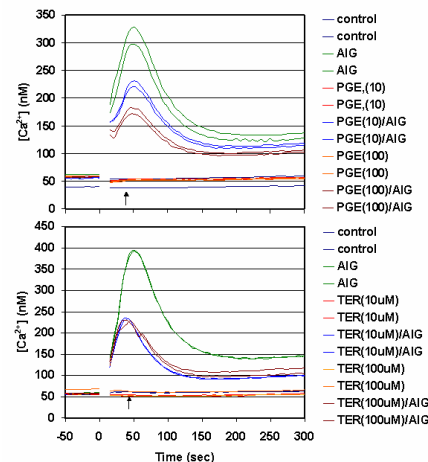
Inputs:

Cells are treated with uniform stimulus. There are 25 ligands applied individually and in pairs

Outputs:



Gene Expression



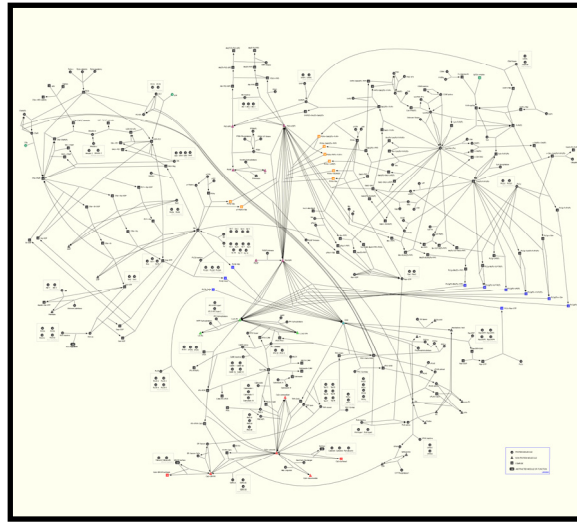
Ca^{2+} , CAMP
Time traces

Other data:

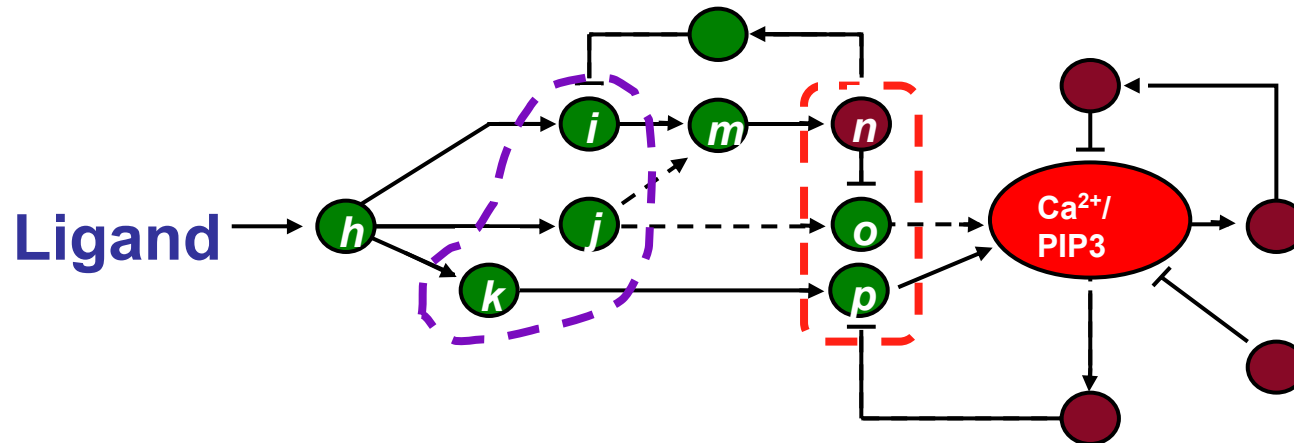
- Protein-Protein interactions
- Phosphorylation states
- All can be repeated with knockdowns

Two efforts: large connection maps and evaluation of minimal models

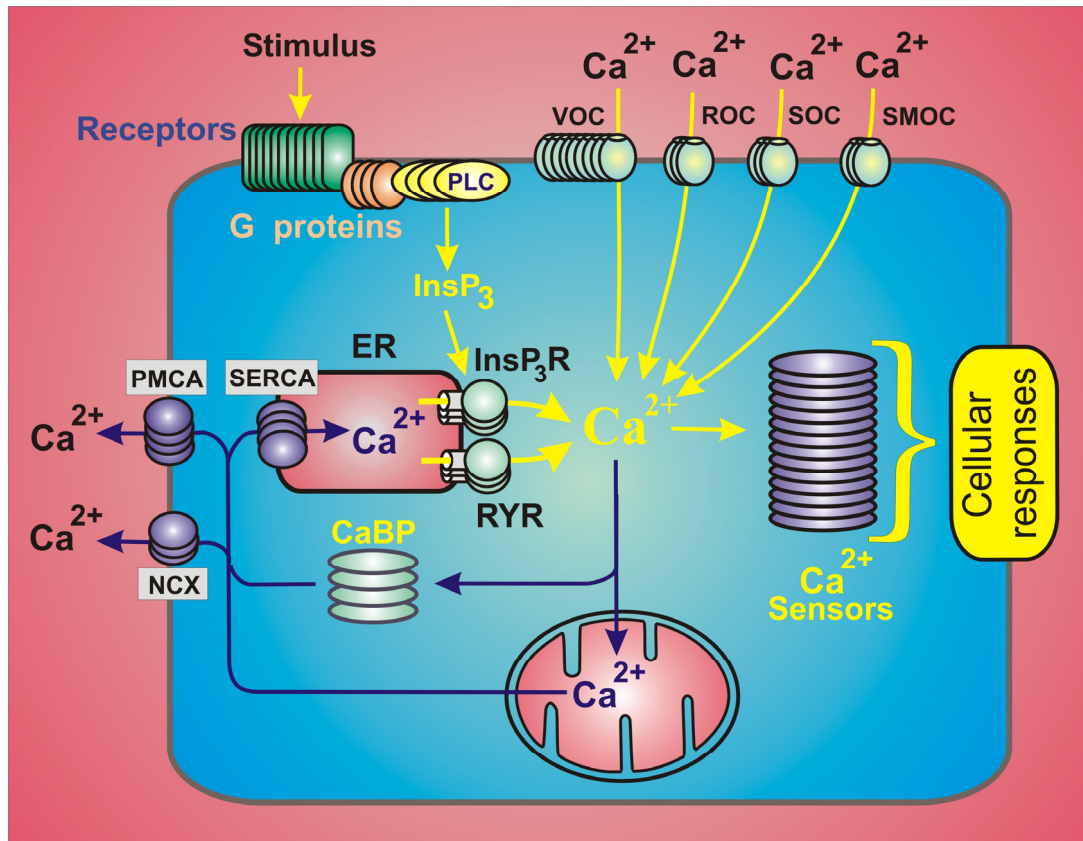
Detailed structure, no kinetics



FXM project: Smaller chunk with Kinetics.



More about Ca^{2+} signaling

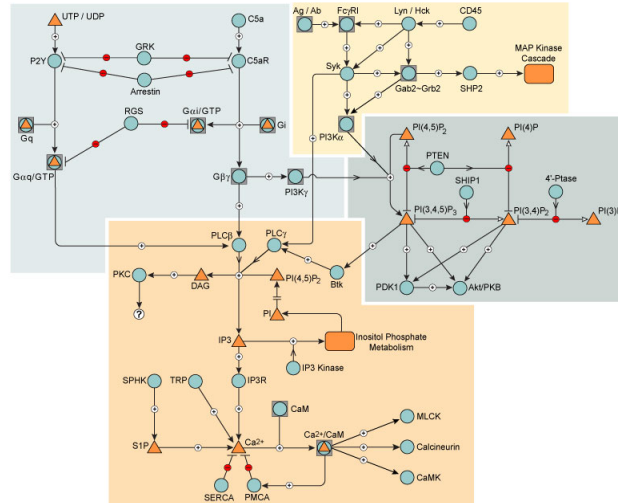


Calcium regulates many responses:

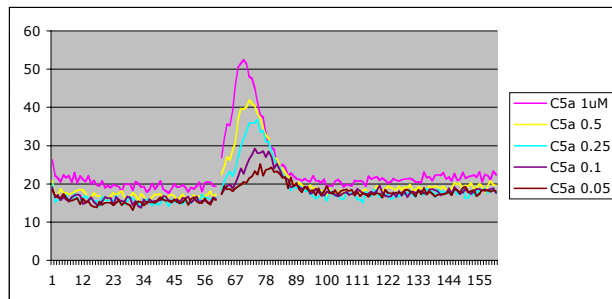
- Synaptic signaling (microseconds)
- Muscle Contraction ($\sim$ milliseconds)
- Gene transcription (hours)
- Embryonic development (days)

AfCS FXM Ca^{2+} data and RNAi Knockdowns

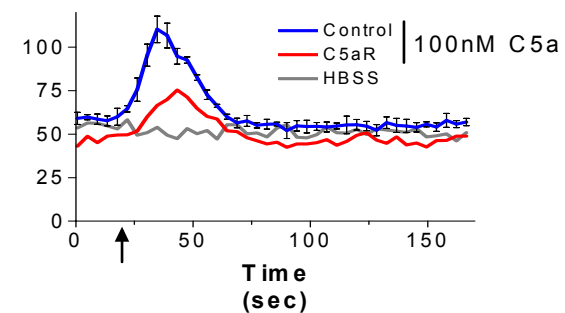
Chunk of pathway with knockdown points (red dots)



Dose Response

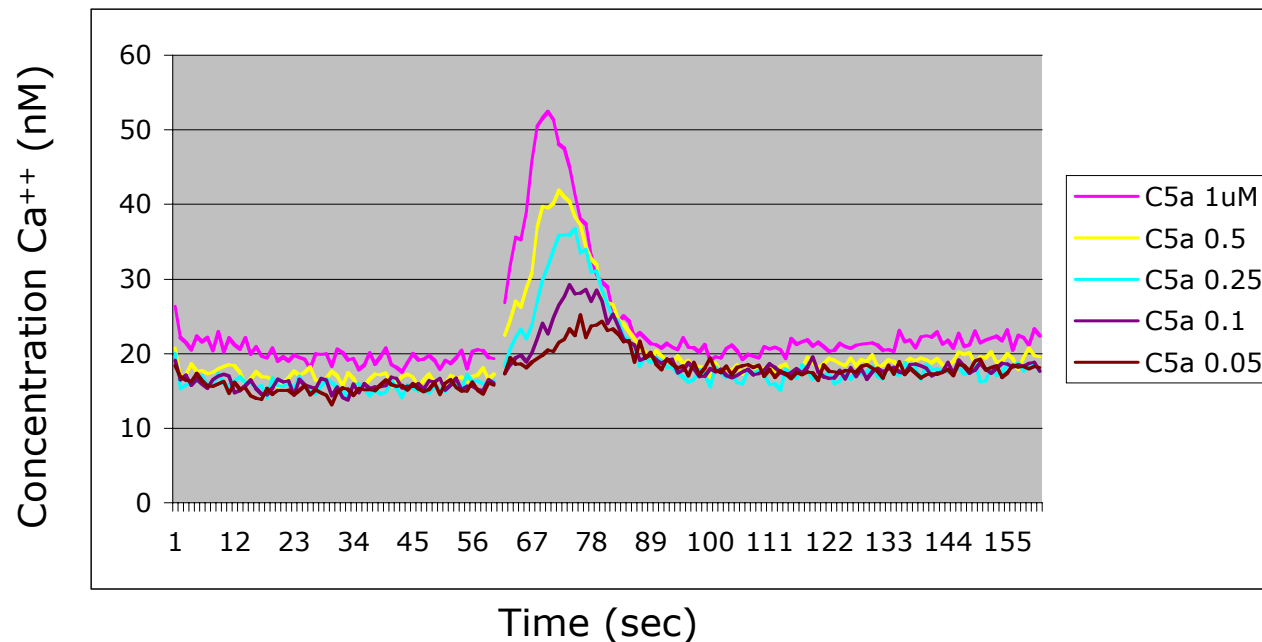


Receptor knockdown



Models, Data, and Uncertainty

- We've examined two Models:
 - A. Goldbeter, Proc. Natl. Acad Sci. 1990
 - T. Wiesner, Modeling in Physiology. 1996
- Today we will focus on the Wiesner model
- Ligand(c5a) dosage data from AFCS



Scalar-valued Data Features

- Considered Ca^{++} response to 4 different ligand dosages:
 - 1000nM
 - 500nM
 - 250nM
 - 100nM
 - From each time-series we extracted 4 scalar-valued features:
 - 95% rise time (sec)
 - Time to peak (sec)
 - Decay time (relative to peak time) to settle to within 5% of peak (sec)
 - Peak concentration relative to steady-state level (μM)
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Extracting data from time series

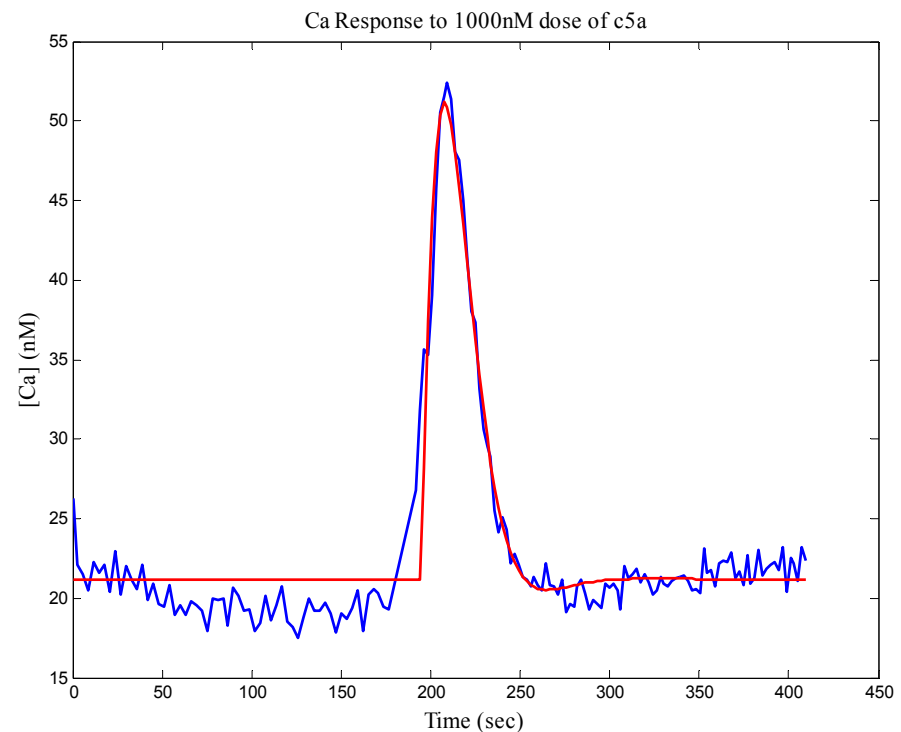
Fit a equation of the following form to the time series data to effectively filter the data in a manner that preserved the character of the spike

$$f(K, \omega, \zeta, \tau, \alpha) = K \frac{\omega}{\sqrt{1 - \zeta^2}} e^{-\zeta \omega * (t - \tau)} \sin(\omega \sqrt{1 - \zeta^2} * (t - \tau)) + \alpha$$

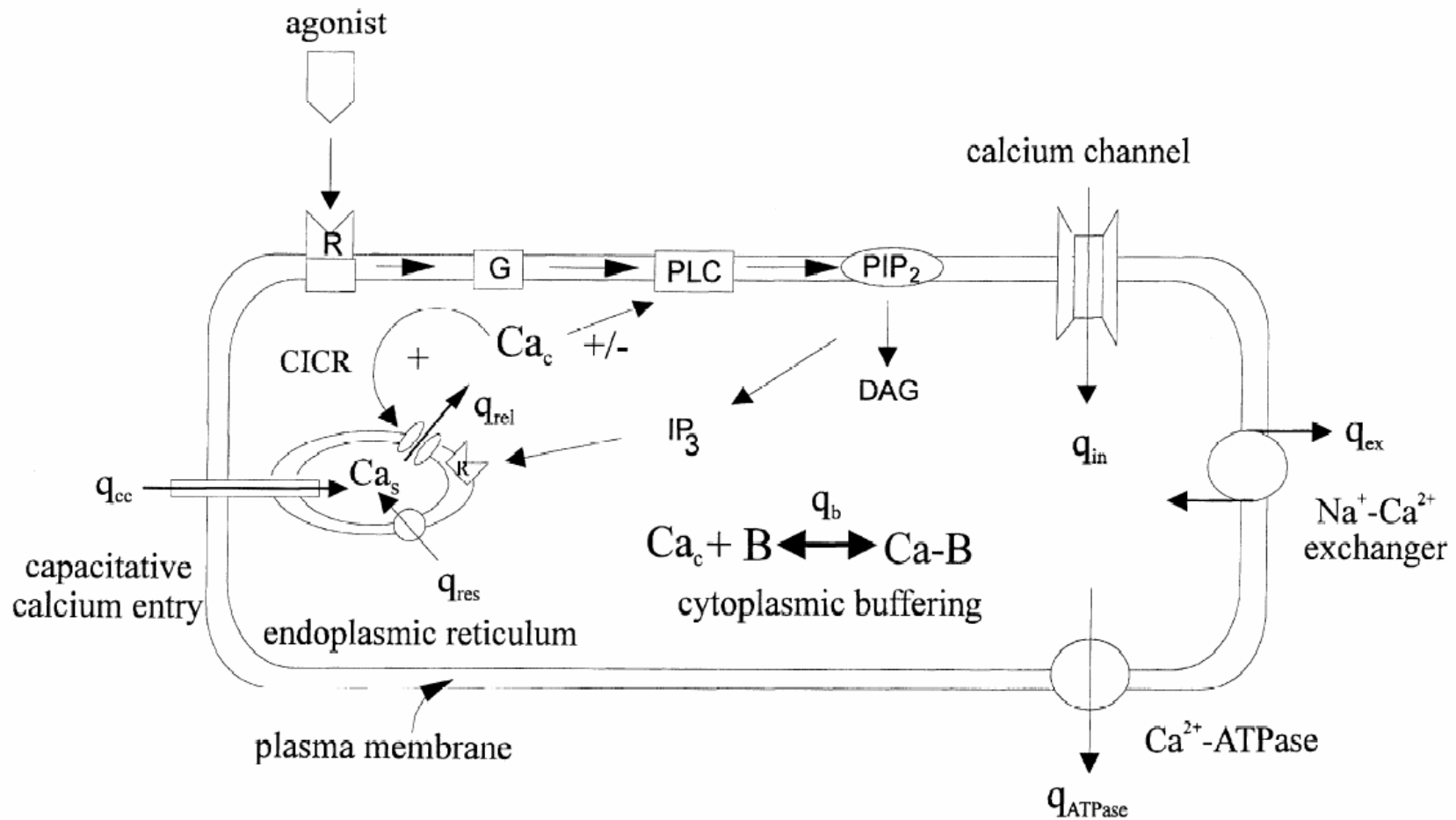
Feature values obtained from f were used as the “data” d_i in our analysis.

For lack of documentation, we specified an uncertainty level $u_i = 0.2d_i$

It would be interesting to use K , ω , ζ , etc. as the features



Schematic of Wiesner Model

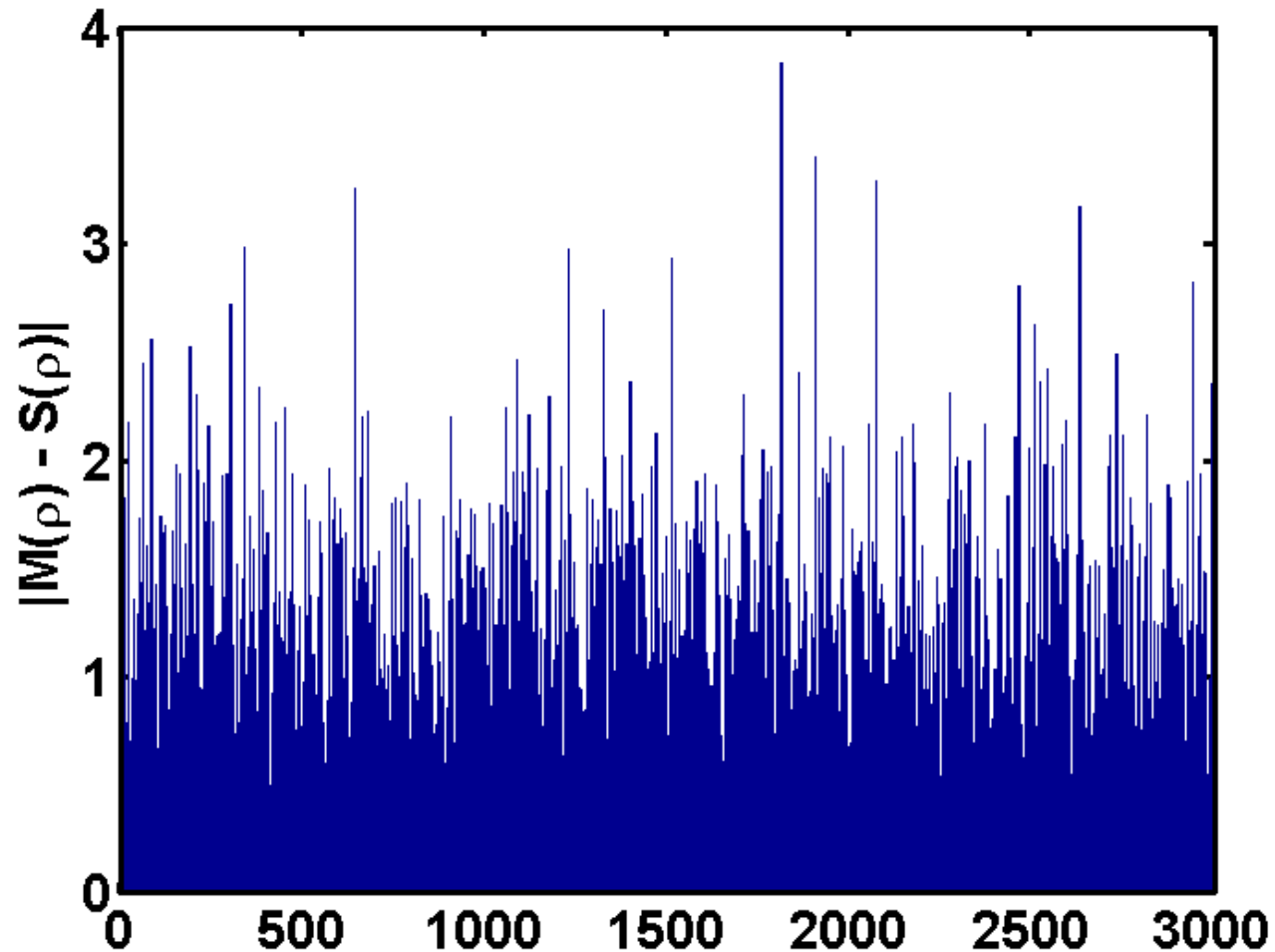


Parameters

- 27 Parameters drawn from
 - Rate constants
 - Cell features (e.g., # of receptors)
 - Initial conditions of species concentration
 - Binding coefficients
 - Papers/literature provided nominal parameter values.
 - We arbitrarily specified the parameter uncertainty level.
 - Typically an order of magnitude for rate constants.
 - Same or less for other parameter types.
 - Parameters were normalized to lie in the hypercube $[-1,1]^n$
 - 16 variables are relevant; the remaining 11 parameters have negligible influence (over the examined ranges) on the model predictions for the chosen features.
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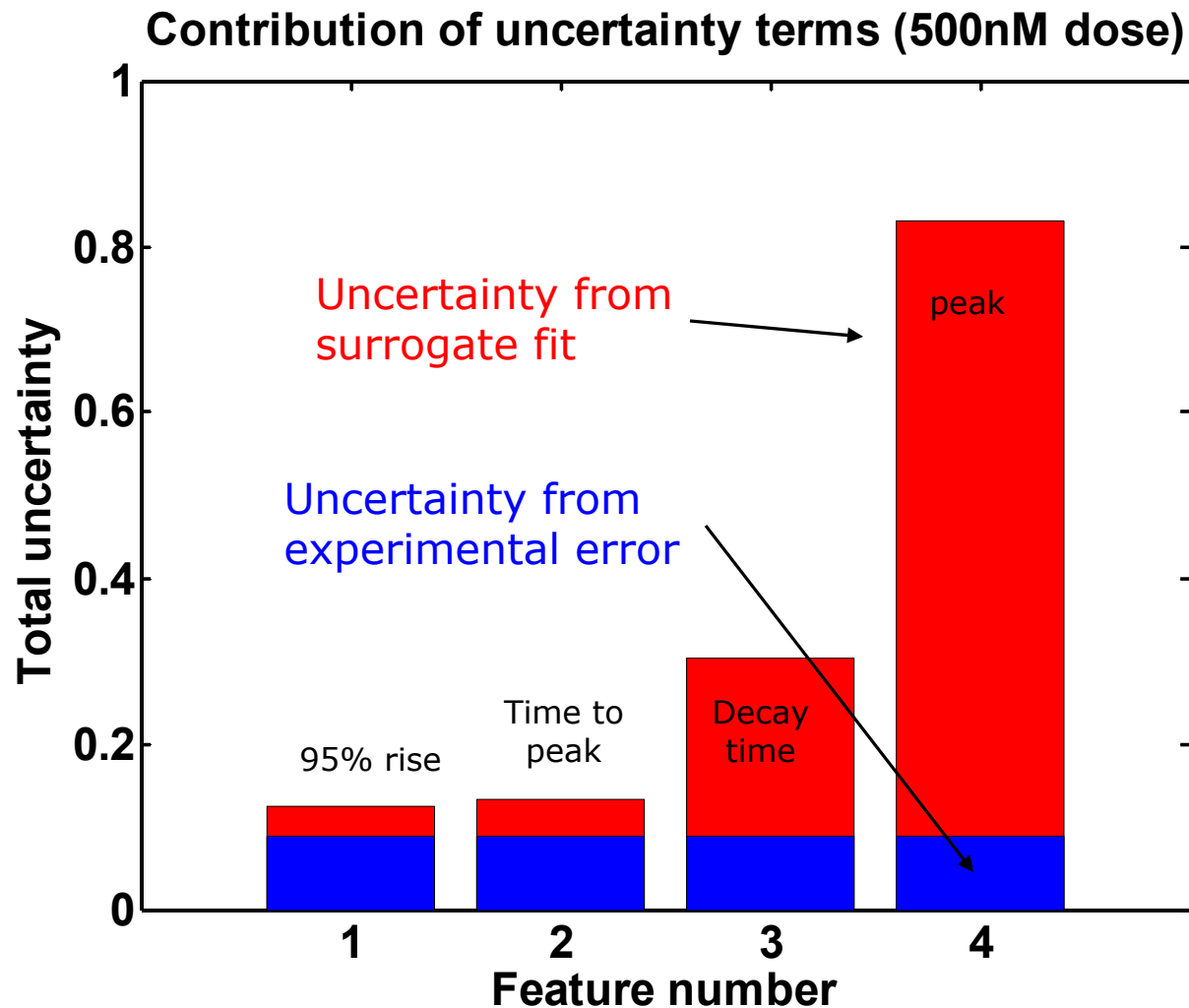
Surrogate Fitting Error Example

Fitting error for peak time at 500nM ligand dose



Surrogate Fits – final analysis

- Large fitting errors in peak Ca^{++} concentration



Consistency analysis

- Question: Does there exist a vector $\rho \in H$ so all feature observations are predicted within their respective uncertainty ranges?

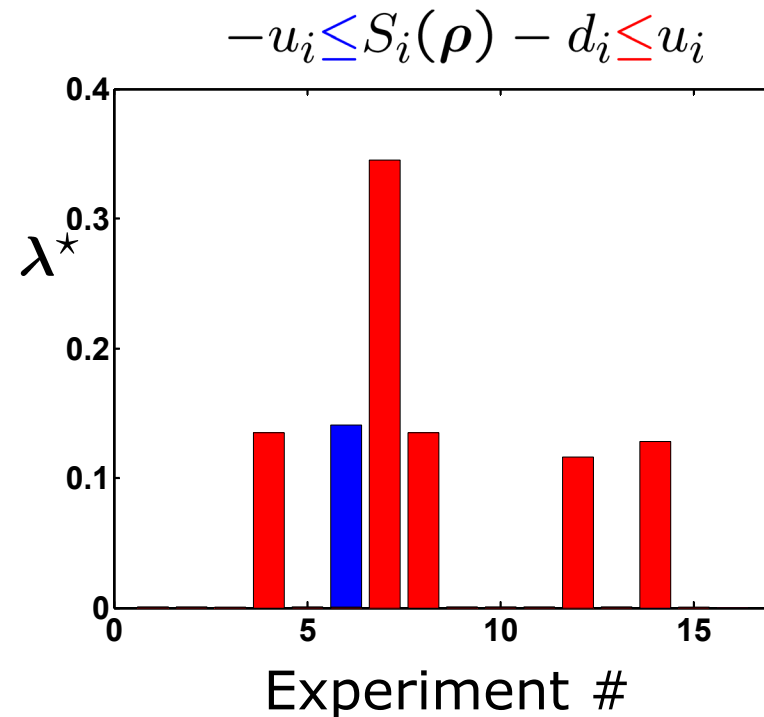
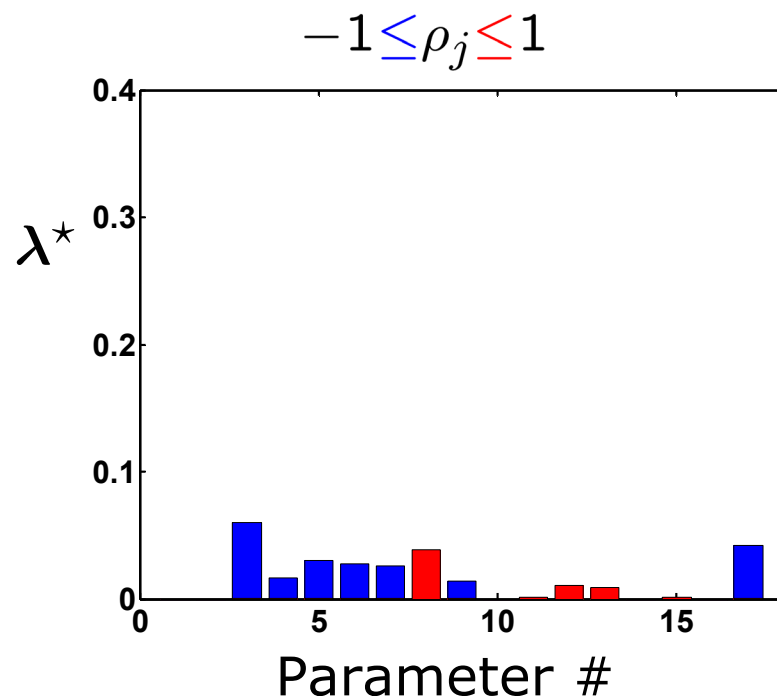
- Consider

$$\begin{aligned} \mathcal{C} = & \text{maximum value of } \gamma \\ \text{subject to the constraints: } & \begin{cases} |\rho_j| \leq 1 & j = 1, \dots, n \\ |S_i(\rho) - d_i| \leq u_i - \gamma & i = 1, \dots, m. \end{cases} \end{aligned}$$

- Negative $\mathcal{C}$ indicates there is no such vector so our assertions regarding the system are inconsistent.
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Consistency Analysis Con't

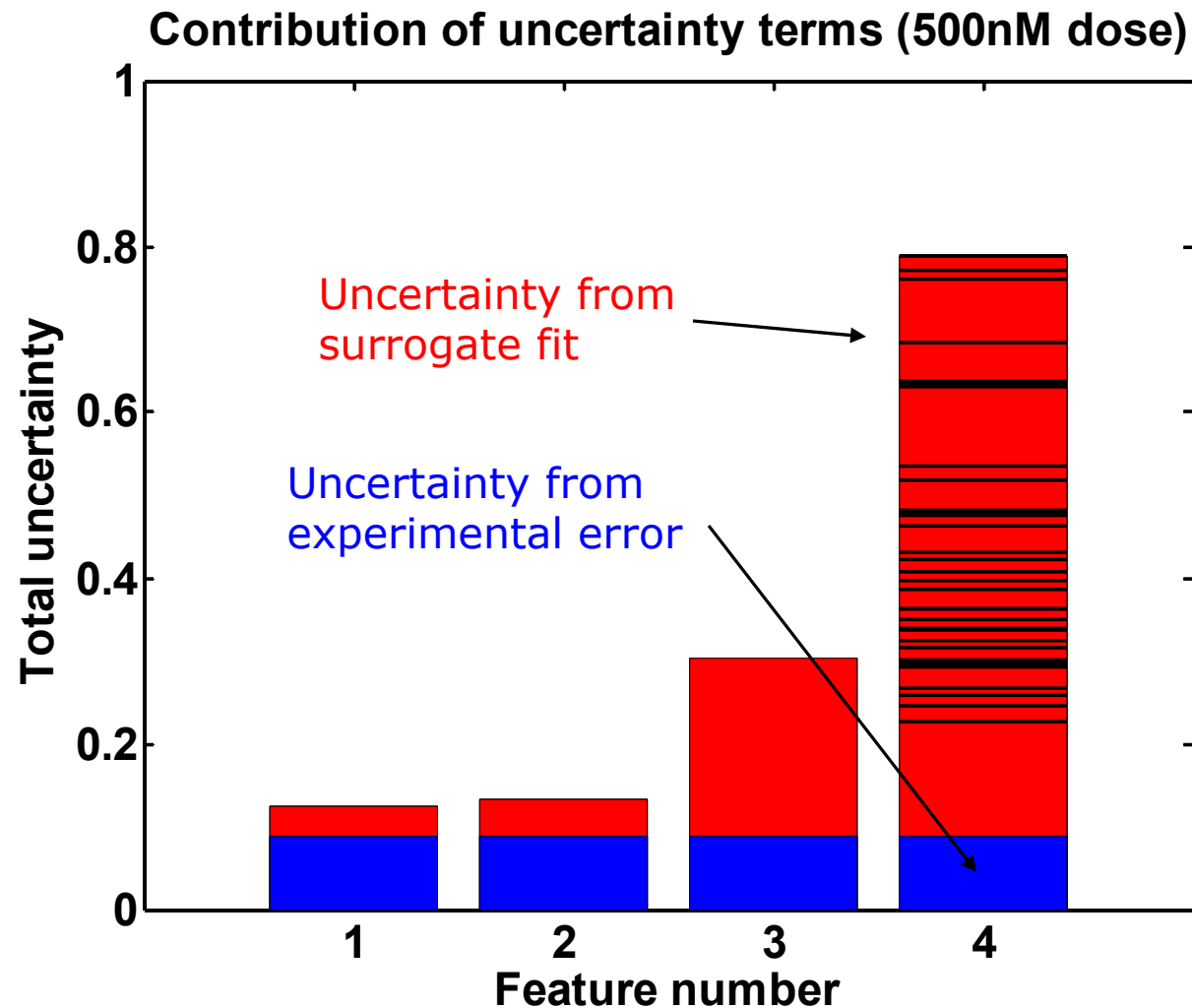
- Including the fitting error in the analysis, our consistency test returns inconclusive results: C 2 [-0.07 0.10]



- Subdivide the cube, re-compute surrogate models and associated fitting errors for the peak concentration models
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Partition and Recompute

Still large fitting errors on a few partitions



But it worked...

- In the worst of the 32 cases, C_2 [-0.15 -0.03]
 - In this case, the consistency measure was largest (but still negative) on the 26th cube.
 - This was not the cube with the worst surrogate fitting errors
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